# Harnessing Platform Methods for mRNA Quality Control: Challenges, Solutions, and Future Perspectives.

#### **USP mRNA Virtual Summit**

Joining Forces to Advance the Quality of mRNA Therapeutics March 11-12, 2025



Mohamad Toutounji, Ph. D - Molgenium -

# Why mRNA?

- Industry growth: \$50B+ market by 2030 (roots in COVID-19 vaccines).
- Therapeutic expansion: Oncology, gene therapy, personalized medicine.
- QC as the Bottleneck: Scalability demands platform approaches.
- Roadmap: Challenges Platform Solutions Future-ready QC.



# Key Challenges in mRNA QC

1. Raw Material Variability

Lipid sources: Lipid batches from different vendors altered LNP size.

Impact: Failed specs, costly delays.

2. Analytical Gaps

Integrity: Ribogreen/qPCR lacks resolution for fragmented mRNA.

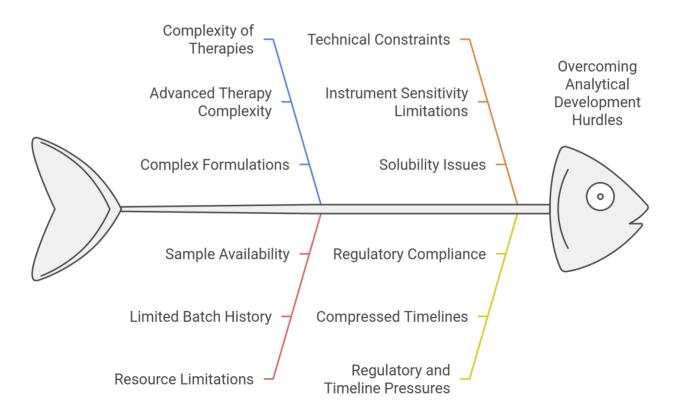
dsRNA: ELISA false positives/positives due to antibody cross-reactivity and assay range.

Poly(A): NGS bias in tail length quantification.



# Key Challenges in mRNA QC: Analytical Methods

## Challenges in Developing Analytical Platform Methods



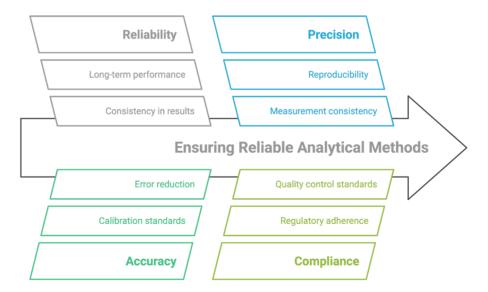


# Method Performance Expectations

### **Defining Method Performance Expectations**

### Purpose

 To validate that analytical methods consistently produce data that are reliable and conform to the predefined performance.





# Method Performance Expectations

### Performance Metrics in Development





### What is Fit-to-Platform Assessment?

#### Definition

Evaluating the suitability of a new analytical method or process to a pre-established platform technology.

Ensures the method aligns with platform standards for performance, compatibility, and reproducibility.

**Key Goals** 

Determine if the method meets platform expectations (e.g., sensitivity, accuracy).

Optimize the platform's capacity to address product-specific challenges.

Significance

Reduces development time by leveraging prior platform knowledge.

Streamlines validation by focusing on critical gaps or enhancements.



### <u>Fit-to-Platform Assessment – Key Components</u>

### Alignment with Platform Capabilities

- Is the new method compatible with the platform's standard design space?
- Does it meet the required performance metrics, including sensitivity, specificity, and robustness?

### Gap Analysis

- Identification of areas where the new method deviates from platform standards.
- Example: Adaptation required for novel analytes or detection targets.



## <u>Fit-to-Platform Assessment – Key Components</u>

#### Risk Assessment

- Quantifying the potential impact of method-platform misalignment on overall product quality.
- Focus on critical quality attributes (CQAs) that could be compromised.

### Optimization

 Adjustments to enhance specificity, linearity, and robustness to meet platform requirements.



### <u>Case Study – Optimizing Specificity in dsRNA Detection</u>

### Background

- dsRNA Detection is a critical step for ensuring mRNA DS safety, as dsRNA is an impurity linked to immune responses.
- ELISA-based assay.

### Challenge

- Standard kits showed cross-reactivity with unrelated nucleic acids, compromising specificity.
- Achieving high specificity while maintaining sensitivity was critical for assessing product safety.

### <u>Case Study – Optimizing Specificity in dsRNA Detection</u>

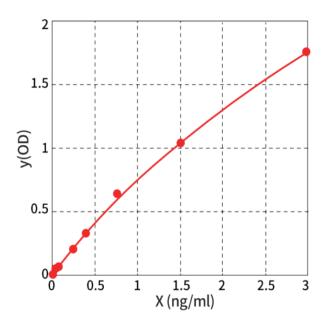
### Approach to Optimization

- 1. dsRNA quantification using ELISA and Fit-to-Platform Evaluation
  - Evaluated the compatibility of the dsRNA detection kit with the existing analytical platform.
  - Identified specificity as the primary limitation through gap analysis.
- 2. Optimizing Specificity
  - Adjusting Kit Design: Modified capture and detection antibodies to reduce crossreactivity.
  - Validation with Controls: Introduced a broader range of negative controls (e.g., single-stranded RNA, DNA) to confirm absence of non-specific binding.

### dsRNA Impurities Control & Detection

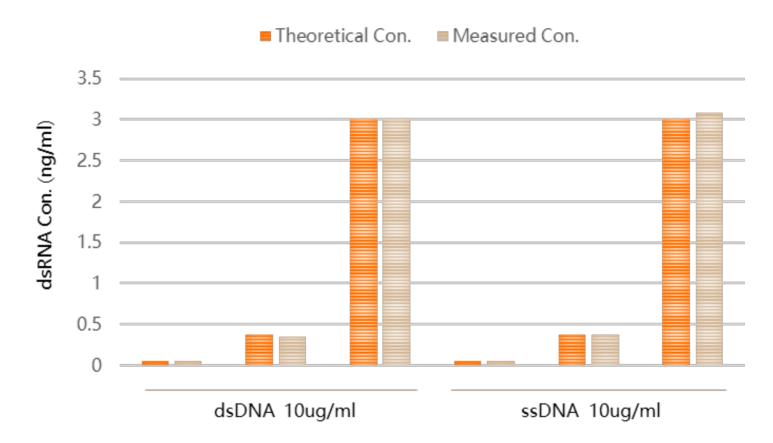
Add 100µl standard/sample per well Add 100µl dsRNA Detection Antibody 37°C, 1h Wash the plate with 1×Wash Buffer at 300µl/well Add 100µl substrate per well Incubate at 37°C for 15min in the dark Add 50µl Stop Reagent per well, and Detect the OD

NovoFast dsRNA ELISA Kit, Cat. No.: RD017



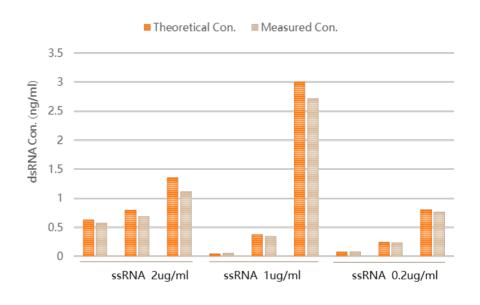
- One-step detection takes only 1.5 hours
- Sensitivity: 0.047ng/ml
- Detection range: 0.047-3ng/ml





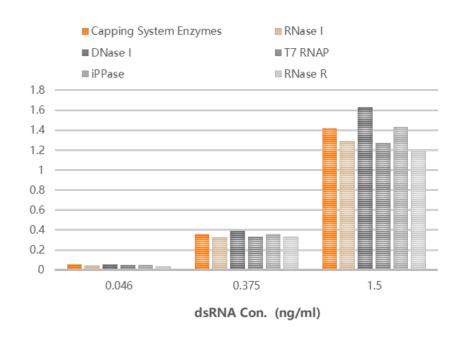


ssRNA μg/ml	Background dsRNA Con. (ng/ml)	dsRNA Con. (ng/ml)	Theoretical dsRNA Con. (ng/ml)	Measured dsRNA Con. (ng/ml)	Recovery%
mRNA1 1019nt 0.2 μg/ml	0.061669	0.000	0.062	0.062	
		0.023	0.085	0.079	6%
		0.188	0.249	0.239	4%
		0.750	0.812	0.764	6%
mRNA 2 2156 nt 1.0 μg/ml	Below detection limit	0.000	Below detection limit		
		0.046	0.046	0.057	24%
		0.375	0.375	0.349	7%
		3.000	3.000	2.715	9%
mRNA3 1019nt 2.0 µg/ml		0.000	0.609	0.609	
	0.609262	0.023	0.632	0.579	8%
		0.188	0.797	0.689	14%
		0.750	1.359	1.112	18%



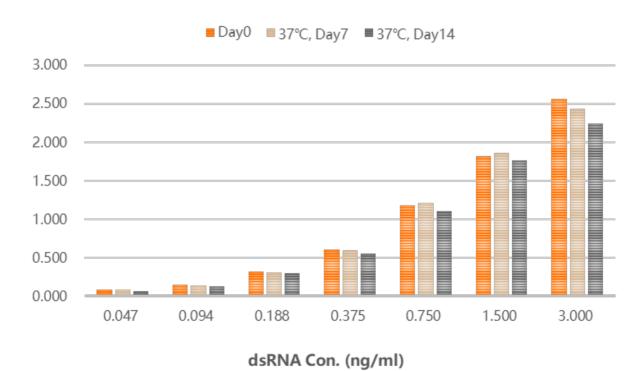


Enzymes (10µg/ml)	Background dsRNA Con. (ng/ml)	dsRNA Con. (ng/ml)	Theoretical dsRNA Con. (ng/ml)	Measured dsRNA Con. (ng/ml)	Recovery%
Capping System Enzymes	Below the detection limit	0.046	0.046	0.055	19%
		0.375	0.375	0.359	4%
		1.500	1.500	1.415	6%
RNase inhibitor	Below the detection limit	0.046	0.046	0.041	11%
		0.375	0.375	0.326	13%
		1.500	1.500	1.286	14%
DNase I	Below the detection limit	0.046	0.046	0.053	16%
		0.375	0.375	0.392	4%
		1.500	1.500	1.628	9%
T7 RNAP	Below the detection	0.046	0.046	0.049	7%
		0.375	0.375	0.333	11%
	iii ii c	1.500	1.500	1.268	15%
iPPase	Below the detection	0.046	0.046	0.051	11%
		0.375	0.375	0.360	4%
	iii ii	1.500	1.500	1.428	5%
RNase R	Delements and the eff	0.046	0.046	0.038	18%
	Below the detection limit	0.375	0.375	0.331	12%
	mint.	1.500	1.500	1.200	20%





### **Accelerated Stability of RD017**





# Design Space and Robustness

#### What is Design Space?

• Defined by ICH Q8 as the multidimensional range of input variables and process parameters that ensure method performance.

#### For dsRNA detection:

 Includes variables such as antibody concentration, reaction time, buffer composition, and temperature.

#### What is Robustness?

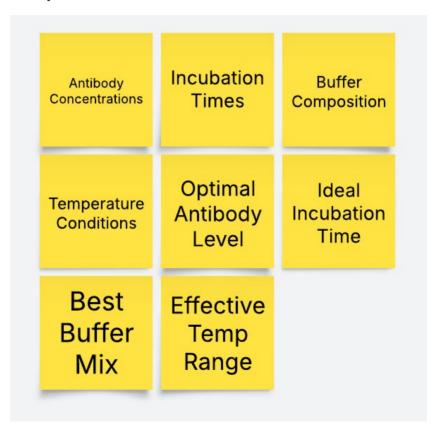
- The method's capacity to remain unaffected by small variations in operating conditions.
- Ensures reliable performance under real-world conditions.



# Design Space and Robustness

Defining the Design Space for dsRNA Detection - Key Variables

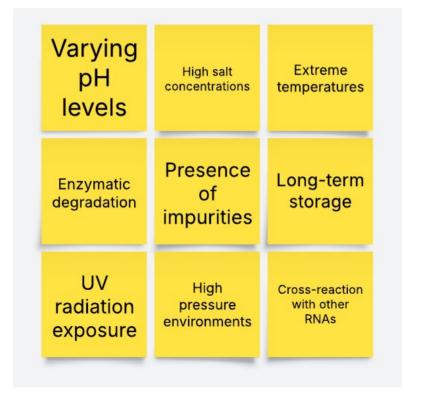
Explored





# Design Space and Robustness

Robustness Testing for dsRNA Detection - Key Stress Conditions Evaluated





#### Definition

 Total Analytical Error (TAE): A statistical measure that combines systematic error (bias) and random error (imprecision) to assess the overall performance of an analytical method.

### Significance

- Provides a holistic view of method reliability.
- Ensures the method delivers results within acceptable accuracy and precision limits.

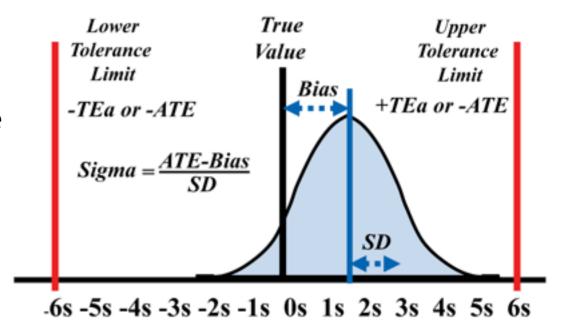
### Components

1. Systematic Error (Bias)

Deviation of the measured value from the true value.

2. Random Error (Imprecision)

Variability observed between repeated measurements.





### Calculating Total Analytical Error

TAE=|Bias|+z×Imprecision (SD)

#### Where:

Bias = Mean of measured values - True value.

z = Standard score for a chosen confidence level (e.g., <math>z = 1.96 for 95%).

SD = Standard deviation of measurements.

#### Interpreting TAE

- Compare TAE to predefined Total Error Allowance (TEA).
- A method is acceptable if:

TAE≤TEA



### Calculating Total Analytical Error for the dsRNA deetction methdo

True Value (spike): 1 ng/mL dsRNA.

Measured Mean: 0.97 ng/mL (Bias = -0.3 ng/mL).

Standard Deviation (SD): 0.25 ng/mL.

TAE Calculation

 $TAE=[-0.3]+(1.96\times0.25)=0.3+0.49=0.79 \text{ ng/mL}.$ 

TEA Benchmark: Defined TEA: ±1 pg/mL.

Conclusion: Since  $\overline{TAE(0.79)} < \overline{TEA(1)}$ , the method is acceptable.



#### Challenges and Best Practices in TAE Evaluation

#### Challenges

- Selecting appropriate TEA limits for complex matrices.
- Balancing sensitivity with precision in low-concentration analytes.
- Addressing variability introduced by operator or equipment changes.

#### **Best Practices**

- Define Clear TEA Limits: Align with product-specific and regulatory requirements.
- Perform Rigorous Testing: Use multiple replicates and conditions to ensure reliability.
- Iterative Optimization: Adjust method parameters to minimize bias and imprecision.



# Leveraging Prior Knowledge for Platform Validation Across Development Stages

#### Role of Prior Knowledge

#### Preclinical/IND Stage

Utilizing prior knowledge to minimize experimental burdens in early stages

#### Phase 1 and Phase 2

Using historical data to optimize validation protocols

# Phase 3/Commercial

Ensuring method reliability through thorough validation or verification





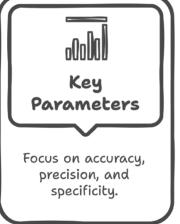




# Regulatory Considerations for Analytical Platforms

### **Expectations for Analytical Platforms**





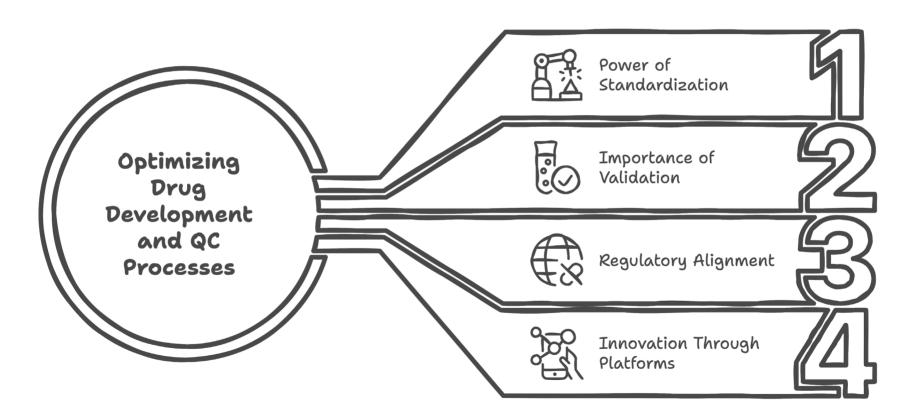








# Take Home Message





# Thank You for Your Attention!



"Together, we pave the way for reliable and innovative drug solutions"